Ectomesenchymal Chondromyxoid Tumor A Neoplasm Characterized by Recurrent RREB1-MKL2 Fusions

Brendan C. Dickson, MD, MSc, *† Cristina R. Antonescu, MD, ‡

Prokopios P. Argyris, DDS, MSc, PhD, § || Elizabeth A. Bilodeau, DMD, MD, ¶

Martin J. Bullock, MD,# Paul D. Freedman, DDS,** Douglas R. Gnepp, MD,††

Richard C. Jordan, DDS, PhD, ‡‡ Ioannis G. Koutlas, DDS, § Cheng-Han Lee, MD, PhD, §§

Iona Leong, BDS, MSc,*[[]] Mihai Merzianu, MD,¶¶ Bibianna M. Purgina, MD,##

Lester D.R. Thompson, MD,*** Bret Wehrli, MD,††† John M. Wright, DDS,‡‡‡

David Swanson, BSc,* Lei Zhang, MD, \ddagger and Justin A. Bishop, MD§§§

Abstract: Ectomesenchymal chondromyxoid tumor is a rare and benign neoplasm with a predilection for the anterior dorsal tongue. Despite morphologic heterogeneity, most cases are characterized by a proliferation of bland spindle cells with a distinctive reticular growth pattern and myxoid stroma. The immunophenotype of these neoplasms is likewise variable; most cases express glial fibrillary acid protein and S100 protein, with inconsistent reports of

- From the *Department of Pathology & Laboratory Medicine, Mount, Sinai Hospital; †Departments of Laboratory Medicine and Pathobiology; |||Oral Pathology & Oral Medicine, Faculty of Dentistry, University of Toronto, Toronto; ##Department of Pathology and Laboratory Medicine, Ottawa Hospital, University of Ottawa, Ottawa; †††Department of Pathology and Laboratory Medicine, London Health Sciences Centre, Western University, London, ON; #Department of Pathology, Dalhousie University, Halifax, NS; §BC Cancer Agency, Vancouver, BC, Canada; ‡Department of Pathology, Memorial Sloan Kettering Cancer Center, New York; **Section of Oral Pathology, New York Presbyterian/Queens, Flushing; ¶Roswell Park Cancer Institute, Buffalo, NY; §Division of Oral and Maxillofacial Pathology, School of Dentistry; ||Department of Biochemistry, Molecular Biology and Biophysics, College of Biological Sciences, University of Minnesota, Minneapolis, MN; ¶Department of Diagnostic Sciences, University of Pittsburgh School of Dental Medicine, Pittsburgh, PA; #Department of Orofacial Sciences, Pathology and Radiation Oncology, University of California San Francisco, San Francisco; ***Southern California Permanente Medical Group, Woodland Hills, CA; ††Department of Pathology, Warren Alpert School of Medicine at Brown University (retired), Providence, RI; ###Texas A&M College of Dentistry; and \$\$\$Department of Pathology, UT Southwestern Medical Center, Dallas, TX
- Conflicts of Interest and Source of Funding: Supported in part by P50 CA140146-01 (C.R.A.); P30-CA008748 (C.R.A.); Kristen Ann Carr Foundation (C.R.A.); Cycle for Survival (C.R.A.). The remaining authors have disclosed that they have no significant relationships with, or financial interest in, any commercial companies pertaining to this article.
- Correspondence: Brendan C. Dickson, MD, MSc, Department of Pathology & Laboratory Medicine, Mount Sinai Hospital, 600 University Ave., Suite 6.500.12.5, Toronto, ON, Canada M5G 1×5 (e-mail: brendan. dickson@sinaihealthsystem.ca).
- Supplemental Digital Content is available for this article. Direct URL citations appear in the printed text and are provided in the HTML and PDF versions of this article on the journal's website, www.ajsp. com.

keratin and myoid marker expression. The molecular pathogenesis is poorly understood; however, a subset of cases has been reported to harbor EWSR1 gene rearrangement. Following identification of an RREB1-MKL2 fusion gene by RNA Sequencing in an index patient, a retrospective review of additional cases of ectomesenchymal chondromyxoid tumors was performed to better characterize the clinical, immunohistochemical, and molecular attributes of this neoplasm. A total of 21 cases were included in this series. A marked predisposition for the dorsal tongue was confirmed. Most cases conformed to prior morphologic descriptions; however, hypercellularity, hyalinized stroma, and necrosis were rare attributes not previously emphasized. The neoplastic cells frequently coexpressed glial fibrillary acid protein, S100 protein, keratin, smooth muscle actin, and/or desmin; a single case was found to contain significant myogenin expression. An RREB1-MKL2 fusion product was identified in 19 tumors (90%), a single tumor (5%) had an EWSR1-CREM fusion product, and the remaining case lacked any known fusion gene by RNA Sequencing. The latter 2 cases subtly differed morphologically from many in the cohort. This series illustrates that recurrent RREB1-MKL2 fusions occur in most. perhaps all, cases of ectomesenchymal chondromyxoid tumor.

Key Words: ectomesenchymal chondromyxoid tumor, tongue, gene rearrangement, RREB1, MKL2, EWSR1, CREM

(Am J Surg Pathol 2018;42:1297-1305)

Ectomesenchymal chondromyxoid tumor is a rare mesenchymal neoplasm—with fewer than 100 reported cases in the English literature—of uncertain origin, with a striking predilection for the anterior dorsal tongue.¹ Tumors occur across a broad age range, predominating in early-mid adulthood, and equally affect males and females.² Clinically, lesions typically present as a small (1 to 2 cm) painless mass, often of longstanding duration.² Simple excision is generally curative; however, a minority of cases have been reported to locally recur.^{2,3}

Morphologically distinctive, tumors are lobulated with thin fibrous septa separating bland polygonal-spindle cells arranged in reticular and globoid patterns.^{2,4,5}

Am J Surg Pathol • Volume 42, Number 10, October 2018

www.ajsp.com | 1297

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

The cytoplasm is pale and eosinophilic. The nuclei are small and ovoid, with scattered atypia, hyperchromasia, pseudonuclear inclusions, and binucleation; mitotic activity, if present, is typically sparse.² Interspersed between the cells is prominent chondromyxoid stroma;² however, overt chondroid differentiation is not generally encountered. The immunophenotype varies-most tumors are characterized by expression of glial fibrillary acid protein and S100 protein, with conflicting reports of immunoreactivity for keratin, smooth muscle actin, and CD56.² There is a single report of a case with SOX10 expression.⁶ Tumors are typically negative for CD34, p63, and epithelial membrane antigen. Ultrastructural examination fails to identify desmosomes or condensations of thin filaments, features that might suggest myoepithelial differentiation.² Rearrangement of EWSR1 has been reported in a subset of cases, although a fusion partner has not, to date, been identified.⁷

Following the identification of an *RREBI-MKL2* fusion gene in an index case of ectomesenchymal chondromyxoid tumor, we undertook a retrospective review of additional cases to better characterize the clinical, immunohistochemical, and molecular attributes of this enigmatic neoplasm.

MATERIALS AND METHODS

Case Selection

An *RREB1-MKL2* fusion gene was identified in the index patient (patient 1) following routine diagnostic RNA Sequencing (RNA-Seq). On the basis of this unexpected observation retrospective searches were performed by each of the contributors for cases diagnosed as ectomesenchymal chondromyxoid tumor (2007 to 2018). Each case was reviewed to confirm the diagnosis before the initiation of immunohistochemical and RNA-Seq testing. This study received institutional Research Ethics Board approval.

Immunohistochemistry

Representative formalin-fixed paraffin-embedded tissue blocks were selected for each case, and 4 μ m sections cut onto positively charged slides. Using standard techniques, staining was performed for glial fibrillary acid protein, S100 protein, keratin (AE1/AE3), smooth muscle actin, desmin, and myogenin (Supplementary Table 1, Supplemental Digital Content 1, http://links.lww.com/PAS/A637). Tumor immunoreactivity was scored based on the percentage of positive cells (0: no staining; 1+: <5%; 2+: 5% to 25%; 3+: 26% to 50%; 4+: 51% to 75%; and 5+: 76% to 100%).

RNA Sequencing

Using formalin-fixed, paraffin-embedded tissue, material was obtained from either scrolls (3 to 4 cut at 10 μ m) or by scrapping unstained sections previously cut onto glass slides (4 to 5 cut at 4 μ m). RNA was extracted using the ExpressArt FFPE Clear RNA Ready kit (Amsbio, Cambridge, MA). RNA-Seq libraries were prepared using 20 to 100 ng total RNA with the TruSight RNA Fusion Panel (Illumina, San Diego, CA). Each sample was sequenced with 76 base-pair paired-end reads on an Illumina MiSeq at 8 samples per flow cell (~3 million reads per sample). The results were analyzed using both the STAR and BOWTIE2 aligners, and Manta and JAFFA fusion callers, respectively.^{8,9}

Fluorescence In Situ Hybridization

Fluorescence in situ hybridization (FISH) for *RREB1* and *MKL2* was performed as previously reported in detail.¹⁰ Briefly, custom bacterial artificial chromosome clone probes were designed to flank the target genes based on the UCSC genome browser (http://genome.ucsc.edu), and obtained from BACPAC sources of Children's Hospital of Oakland Research

Patient	Age (y)	Sex	Location	Size and Clinical Findings				
1	31	F	Tongue, NOS	0.9 cm. Excised with negative margin				
2	48	Μ	Tongue, dorsal, tip	0.6 cm. Shave biopsy. No evidence of recurrence at 1 y				
3	39	F	Tongue, left dorsal, posterior	1.1 cm. <2 mo follow-up				
4	54	F	Tongue, dorsal, anterior	1.4 cm. Re-excision, with positive margin. No report of recurrence				
5	54	Μ	Tongue, dorsal	Not available				
6	23	Μ	Tongue, left anterior	3.0 cm. No evidence of recurrence at 2 y				
7	13	F	Tongue, right midline dorsal	2.0 cm soft mass				
8	45	F	Tongue, left dorsal	Size not specified. 6-7 mo asymptomatic mass				
9	54	Μ	Tongue, left anterior	1.2 cm. Excised with negative margins. No evidence of recurrence at 1.4 y				
10	35	Μ	Tongue, left anterior	0.7 cm. Excised with positive margins. Recurred at 41 mo. No further recurrences after 4 y				
11	14	F	Tongue, midline anterior	1.5 cm. Excised with positive margins. No evidence of recurrence after 8.5 y				
12*	49	F	Tongue, dorsal lateral	2.1 cm				
13*	33	F	Tongue, NOS	Not available				
14	50	F	Tongue, dorsal tip	2.4 cm. Excised with negative margin. LTF				
15*	53	F	Tongue, NOS	Not available				
16*	59	Μ	Tongue, NOS	Not available				
17*	31	F	Tongue, NOS	Not available				
18*	51	Μ	Tongue, midline dorsal, anterior	2.5 cm. 15-y history before excision				
19	40	F	Tongue, right dorsal, anterolateral	1.0 cm. Slow-growing lump ×3-5 mo. Margin focally positive, but no report of recurrence				
20	51	Μ	Tongue, NOS	0.7 cm. Margin focally positive, with no report of recurrence				
21	15	F	Tongue, right dorsal, posterior	0.7 cm				

*Previously reported.7,11

F indicates female; LTF, lost to follow-up; M, male; NOS, not otherwise specified.

1298 | www.ajsp.com

Institute (Oakland, CA; http://bacpac.chori.org) (Supplementary Table 2, Supplemental Digital Content 2, http://links. lww.com/PAS/A638). DNA from each bacterial artificial chromosome was isolated and then labeled with fluorochromes by nick translation. Slides were prepared using formalin-fixed, paraffin-embedded tissue cut at 4 μ m. The slides were deparaffinized, pretreated, and then hybridized with the denatured probes. Following an overnight incubation, the slides were rinsed, stained with 4',6-diamidino-2-phenylindole, mounted, and examined using a Zeiss fluorescence microscope (Zeiss Axioplan, Oberkochen, Germany).

RESULTS

A total of 21 cases were identified from the contributing institutions; 6 of the cases (patients 13, 14, 16, 17, 18, 19) have previously been reported.^{7,11} The mean patient age was 40 years (range, 13 to 59 y). There were 13 females and 8 males (ratio, 1.6:1). Each of the tumors arose on the dorsal tongue; most were anterior, and a minority posterior. Most cases were

treated by simple excision. Despite frequent microscopic extension to inked resection margins, only a single case was associated with local recurrence (patient 10) (Table 1).

The size of the tumors ranged from 0.6 to 3.0 cm (mean, 1.5 cm); virtually all of the cases were circumscribed, with a pushing border; several tumors contained focal infiltration into the surrounding skeletal muscle. The overlying squamous epithelium was intact in all cases, lacking any well-developed pseudoepitheliomatous hyperplasia. The tumors were frequently multilobulated; the lobules were either separated by areas of endogenous stroma, or more frequently by fibrous septa (Fig. 1). The cells ranged from polygonal to spindlestellate. The cytoplasm was pale and eosinophilic to lightly basophilic. The nuclei were round to ovoid and frequently lobulated and hyperchromatic. Scattered cells showed atypia, pseudonuclear inclusions, and binucleation. Almost all of the cases lacked conspicuous mitotic activity; however, rare cases contained 1 to 2 mitoses per 10 high-power fields. Similar to prior reports, the architecture included sheets, cords, reticular, and globoid patterns; cellular fascicles, microcystic-cystic, and



FIGURE 1. Representative photomicrographs from a conventional case of ectomesenchymal chondromyxoid tumor (patient 3). A, Low-power magnification showing circumscribed neoplasm with pushing margin. B, Intermediate magnification highlighting the presence of fibrous septa separating tumor lobules. C, Spindle-stellate cells with a reticular, or "net-like" pattern and abundant myxoid stroma. D, Scattered cytologic atypia and multinucleation; however, mitotic activity is inconspicuous.

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

www.ajsp.com | 1299

papillary patterns were rarely encountered. Occasional interstitial hemorrhage and/or hemosiderin deposition—in the absence of prior sampling—was identified. A single case contained necrosis and dystrophic calcification (Fig. 2). The intervening stroma was myxoid and pale and basophilic;

occasionally it was hyalinized; very rarely, isolated foci vaguely suggested hyaline cartilage. Sunburst amianthoid fibers were not identified.

The results of immunohistochemical staining are summarized in Table 2 and Figure 3. Briefly, similar to



FIGURE 2. Less common features observed in ectomesenchymal chondromyxoid tumor. A, Cellular spindle cell areas with a vague fascicular pattern. B, Eosinophilic hyaline stroma. C, Stroma vaguely suggestive of hyaline cartilage. D, Microcystic-cystic pattern. E, Nodular-cribriform pattern. F, Necrosis.

1300 | www.ajsp.com

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

	Immunohistochemistry						Molecular	
Patient	GFAP	S100	Keratin	Desmin	Myogenin	SMA	Method	Result
1	_	3+	_	1+	NP	1+	RNA-Seq	RREB1-MKL2
2	-	-	-	-	-	-	RNA-Seq	EWSR1-CREM
3	5+	5+	1+	1+	-	3+	RNA-Seq	RREB1-MKL2
4	4+	3+	1+	-	-	1+	RNA-Seq	Negative
5	NP	NP	NP	NP	NP	NP	RNA-Seq	RREB1-MKL2
6	5+	5+	2+	1+	2+	2+	RNA-Seq	RREB1-MKL2
7	5+	2+	-	NP	NP	-	RNA-Seq	RREB1-MKL2
8	5+	3+	-	NP	NP	NP	RNA-Seq	RREB1-MKL2
9	1+	3+	-	-	-	3+	RNA-Seq	RREB1-MKL2
10	2+	3+	-	-	-	3+	RNA-Seq and FISH	RREB1-MKL2
11	5+	3+	2+	4+	-	2+	RNA-Seq	RREB1-MKL2
12	4+	2+	1+	1+	-	NP	FISH	RREB1-MKL2
13	5+	2+	1+	1+	-	NP	FISH	RREB1-MKL2
14	4+	1+	-	1+	-	NP	FISH	RREB1-MKL2
15	4+	NP	-	NP	-	NP	FISH	RREB1-MKL2
16	4+	1+	-	-	-	NP	FISH	RREB1-MKL2
17	2+	NP	-	NP	-	NP	FISH	RREB1-MKL2
18	5+	5+	2+	1+	-	3+	FISH	RREB1-MKL2
19	5+	2+	2+	1+	-	NP	FISH	RREB1-MKL2
20	4+	1+	1+	3+	1+	3+	FISH	RREB1-MKL2
21	5+	2+	2+	-	-	_	FISH	RREB1-MKL2
- indica	tes negative; GF	AP, glial fibrill	ary acid protein;	NP, not performe	d; SMA, smooth mu	scle actin.		

TABLE 2. Summary of Immunohistochemical and Molecular Attributes of Ectomesenchymal Chondromyxoid Tumor

prior reports, there was relatively consistent expression of glial fibrillary acid protein. Immunoreactivity for S100 protein, pancytokeratin, smooth muscle actin, and desmin expression was common, but tended to be weak and/or focal. A single case contained significant myogenin expression. Immunohistochemistry was performed for SOX10 (N=7) and p63 (N=2), which were negative (data not shown).

Following RNA-Seq the index patient was found to have an RREB1-MKL2 fusion gene. RNA-Seq was subsequently performed on patients 2 to 11. As a mean of independent verification of the RNA-Seq fusion product, FISH testing was applied to the recurrent lesion of patient 10. The remaining cases (patients 12 to 21) were examined by FISH alone (Fig. 4). All but 2 cases (90.5%) were found to harbor RREB1-MKL2 fusion genes. One patient was found to have an EWSR1-CREM fusion gene (patient 2), while the remaining case lacked any fusion candidates, despite suitable RNA (patient 4) (Fig. 5). RNA-Seq of the cases with an RREB1-MKL2 fusion product demonstrated identical breakpoints involving exon 8 of RREB1 (NCBI Reference Sequence: NM_001003699.3) and exon 11 of MKL2 (NM_001308142.1). The single case with an EWSR1-CREM fusion had breakpoints involving exon 8 of EWSR1 (NM_013986.3), and either exon 7 or 3 of CREM (NM 181571.2, or NM 001352467.1) depending on whether CREM transcript variant 1 or 32.

DISCUSSION

Ectomesenchymal chondromyxoid tumor is a rare mesenchymal neoplasm of unknown histogenesis. Our series, among the largest to date, confirms prior reports of the relatively narrow clinical distribution of this enigmatic tumor; in addition, we expand the morphologic and immunohistochemical spectrum of this neoplasm. Finally, we demonstrate that the majority of ectomesenchymal chondromyxoid tumors are characterized by *RREB1-MKL2* fusions.

In this series tumors were found in patients from across a relatively broad age range (13 to 59 y), with a slight female predilection. All of our cases originated in the tongue. Indeed, virtually all prior reports of ectomesenchymal chondromyxoid tumor have occurred in this location. While a hard palate origin has been suggested in 2 reports,^{12,13} the first has been previously questioned,^{3,14–16} and the second is also noted to lack a prototypic morphology and immunophenotype. Most of the tumors in our series arose in the anterior tongue, but a minority involved the base of tongue.^{4,17,18} Where clinically specified, all cases were dorsally situated. Despite several cases in this series with positive margins following excision, only a single case locally recurred and none metastasized. This would appear to confirm a clinically benign behavior.

Ectomesenchymal chondromyxoid tumor has a distinct morphology and immunophenotype. Since its initial report by Smith et al,² there have been few revisions to the original comprehensive histopathologic description; indeed, our findings were largely similar. The cells ranged from polygonal to spindle and stellate. Most cases contained, at least focally, the prototypic "net-like" (reticular) arrangement of cells with abundant myxoid stroma. Clefting, papillary, cystic, and more solid growth patterns were occasionally observed. Hyalinized eosinophilic stroma, calcification, hypercellularity, and necrosis were rare events. Similar to the earlier reports, immunoreactivity for glial fibrillary acid protein, S100 protein, keratin, and actin were frequently observed;² glial fibrillary protein notwithstanding, immunoreactivity tended to be patchy and/or weak. While there are conflicting reports on desmin

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

www.ajsp.com | 1301



FIGURE 3. Representative photomicrographs of immunophenotype of ectomesenchymal chondromyxoid tumor (patient 3). Glial fibrillary acid protein (A), S100 protein (B), keratin (AE1/AE3) (C), desmin (D), smooth muscle actin (E), and myogenin (F).

expression,^{18–20} our results suggest this is relatively common, although focal. We observed significant nuclear myogenin expression in a single, otherwise unremarkable, case.

Until now the molecular pathogenesis of ectomesenchymal chondromyxoid tumor has remained elusive. In a series of 11 cases Argyris and colleagues identified *EWSR1* rearrangement in 3 cases (27%); 2 subsequent cases tested by Laco et al⁶ failed to show *EWSR1* rearrangement. Following the discovery of a *RREB1-MKL2* fusion gene in our index patient, we confirmed, using a combination of RNA-Seq and FISH, this fusion product

1302 | www.ajsp.com

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.



FIGURE 4. Representative FISH in ectomesenchymal chondromyxoid tumor (patient 20). Rearrangement of: *MKL2* (A) and *RREB1* (B) (both images: red, centromeric; green, telomeric).



FIGURE 5. Representative photomicrographs of ectomesenchymal chondromyxoid tumor lacking *RREB1-MKL2* fusion partners. A and B, A neoplasm with an *EWSR1-CREM* fusion product contains lobules of bland cells. C and D, A neoplasm lacking any gene fusions by RNA-Seq. The tumor is infiltrative and contains an adipocytic component. The lesion is cellular with less conspicuous myxoid component.

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

in 19 of 21 cases (90%). The 2 cases lacking an RREB1-MKL2 fusion did not possess an archetypical morphology. Nevertheless, as they currently fall within the accepted clinical and pathologic spectrum of ectomesenchymal chondromyxoid tumor, there is currently insufficient evidence to warrant rejection of the original diagnosis. One case (patient 4) was cellular with infiltrative tongues resembling a somewhat atypical case presented by Portnof et al.³ The second case (patient 2) was found to have an EWSR1-CREM fusion product by RNA-Seq; this case contained small nodules of polygonal cells with indistinct cell borders and prominent slit-like spaces. This was morphologically similar to at least 2 of the prior cases of ectomesenchymal chondromyxoid tumor with EWSR1 rearrangement.⁷ While further study is required, it is possible that cases with EWSR1 rearrangement in this context may ultimately be more appropriately classified under the recently characterized family of myxoid mesenchymal tumors with EWSR1-CREB family gene fusions.¹⁰ In contrast to that seminal report, our patient was considerably older than the patients in that series, and our tumor was extracranial. The limited literature on myxoid mesenchymal neoplasms with EWSR1-CREM makes it difficult to draw meaningful inferences to our case; moreover, one of us (B.C.D.) has recently encountered a malignant small round cell neoplasm with an *EWSR1-CREM* fusion gene in an extracranial location.

Interestingly, a recent case of so-called 'biphenotypic "oropharyngeal" sarcoma' was reported to possess an *RREB1-MKL2* gene fusion.²¹ In their article, Siegfried and colleagues describe a 3.5 cm parapharyngeal mass comprised of fascicles of bland spindle-ovoid cells with uniform nuclei and minimal mitotic activity, and occasional pseudoangiectatic and cystic spaces; immunohistochemistry demonstrated patchy staining for S100 protein, desmin, smooth muscle actin, and myogenin. Anatomic proximity notwithstanding, this case does not appear to involve the base of tongue. And, while some of the cases in our series contained areas of increased cellularity and a fascicularherringbone pattern, this was not a dominant finding.²¹ Whether biphenotypic "oropharyngeal" sarcoma represents an atypical example of ectomesenchymal chondromyxoid tumor, or a distinct entity with a pleiotropic fusion geneanalogous to the relationship of EWSR1-ATF1 in clear cell sarcoma, angiomatoid fibrous histiocytoma and hyalinizing clear cell carcinoma of salivary gland—is presently unclear.

The histogenesis of ectomesenchymal chondromyxoid tumor has been a subject of debate. It has been suggested to originate from an uncommitted ectomesenchymal cell derived from the neural crest.^{1,2} Alternatively, it has been proposed to represent a myoepithelial-derived tumor of either minor salivary glands or soft tissue.^{2,22} Our molecular findings, combined with the unique clinical and histopathologic attributes of this neoplasm, suggest ectomesenchymal chondromyxoid tumor represents a distinct entity. *MKL2* encodes Myocardin Like 2—a transcription coactivator of serum response factor—that is involved in smooth and skeletal muscle differentiation,^{23,24} and neuronal development.²⁵ Siegfried et al²¹ eloquently describe how the *RREB1-MKL2*

fusion gene—which is associated with increased *MKL2* expression—accounts for a biphenotypic immunophenotype. There is a precedent for *MKL2* fusions in another mesenchymal neoplasm; *C11orf95-MKL2* fusions occur in chondroid lipoma^{26,27} which, similar to ectomesenchymal chondromyxoid tumor, is variably characterized by myxoid-chondroid matrix. Similar to many other translocation-associated neoplasms that undergo fundamental molecular re-reprogramming²⁸—with a concomitant absence of an overt line of differentiation (eg, Ewing sarcoma and synovial sarcoma)—it is conceivable that a cell of origin may remain elusive for some time; that being said, Smith et al.'s² original hypothesis of an uncommitted ectomesenchymal cell progenitor remains a distinct possibility.

In summary, this study represents the largest series of molecularly interrogated ectomesenchymal chondromyxoid tumors to date. We confirm the strong predilection of this neoplasm for the dorsal aspect of the tongue, and expand the morphologic and immunohistochemical spectrum of findings for this tumor. Finally, we demonstrate that the overwhelming majority—if not all, if 2 morphologic outliers can be excluded—of cases of ectomesenchymal chondromyxoid tumor are characterized by an *RREB1-MKL2* fusion gene.

ACKNOWLEDGMENTS

The authors are grateful to Anthony Wing and Jasmine Wong for their expertise in immunohistochemistry; Grace Murray (Illumina, San Diego, CA) for generously providing test kits; and, Evangeline Agro and Sharon Crafter for facilitating RNA-Seq testing.

REFERENCES

- Bishop JA, Gnepp DR, Ro JY. Ectomesenchymal chondromyxoid tumour. In: El-Naggar AK, Chan JKC, Grandis JR, Takata T, Slootweg PJ, eds. WHO Classification of Head and Neck Tumours. Lyon: IARC; 2017:119–120.
- Smith BC, Ellis GL, Meis-Kindblom JM, et al. Ectomesenchymal chondromyxoid tumor of the anterior tongue. Nineteen cases of a new clinicopathologic entity. *Am J Surg Pathol.* 1995;19:519–530.
- 3. Portnof JE, Friedman JM, Reich R, et al. Oral ectomesenchymal chondromyxoid tumor: case report and literature review. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2009;108:e20–e24.
- 4. Seo SH, Shin DH, Kang HJ, et al. Reticulated myxoid tumor of the tongue: 2 cases supporting an expanded clinical and immunopheno-typic spectrum of ectomesenchymal chondromyxoid tumor of the tongue. *Am J Dermatopathol.* 2010;32:660–664.
- Ide F, Mishima K, Saito I. Ectomesenchymal chondromyxoid tumor of the anterior tongue with myxoglobulosislike change. *Virchows Arch.* 2003;442:302–303.
- Laco J, Mottl R, Hobling W, et al. Cyclin D1 expression in ectomesenchymal chondromyxoid tumor of the anterior tongue. *Int J Surg Pathol.* 2016;24:586–594.
- Argyris PP, Bilodeau EA, Yancoskie AE, et al. A subset of ectomesenchymal chondromyxoid tumours of the tongue show EWSR1 rearrangements and are genetically linked to soft tissue myoepithelial neoplasms: a study of 11 cases. *Histopathology*. 2016; 69:607–613.
- Liu S, Tsai WH, Ding Y, et al. Comprehensive evaluation of fusion transcript detection algorithms and a meta-caller to combine top performing methods in paired-end RNA-seq data. *Nucleic Acids Res.* 2016;44:e47.
- 9. Chen X, Schulz-Trieglaff O, Shaw R, et al. Manta: rapid detection of structural variants and indels for germline and cancer sequencing applications. *Bioinformatics*. 2016;32:1220–1222.

1304 | www.ajsp.com

Copyright © 2018 Wolters Kluwer Health, Inc. All rights reserved.

- Kao YC, Sung YS, Zhang L, et al. EWSR1 fusions with CREB family transcription factors define a novel myxoid mesenchymal tumor with predilection for intracranial location. *Am J Surg Pathol.* 2017;41:482–490.
- Schep LA, Bullock MJ, Taylor SM. Ectomesenchymal chondromyxoid tumour of the dorsal tongue presenting with impaired speech. *Case Rep Otolaryngol.* 2016;2016:7342910.
- Nigam S, Dhingra KK, Gulati A. Ectomesenchymal chondromyxoid tumor of the hard palate–a case report. J Oral Pathol Med. 2006;35: 126–128.
- Gouvea AF, Diaz KP, Leon JE, et al. Nodular lesion in the anterior hard palate. Oral Surg Oral Med Oral Pathol Oral Radiol. 2012;114:154–159.
- Kato MG, Erkul E, Brewer KS, et al. Clinical features of ectomesenchymal chondromyxoid tumors: a systematic review of the literature. *Oral Oncol.* 2017;67:192–197.
- Ide F. Chondromyxoid tumor of palate. J Oral Pathol Med. 2006;35: 523–524; author reply 524.
- 16. Allen CM. The ectomesenchymal chondromyxoid tumor: a review. *Oral Dis.* 2008;14:390–395.
- Cardin MJ, Fiset PO, Zeitouni AG, et al. Ectomesenchymal chondromyxoid tumour of the posterior tongue. *Head Neck Pathol.* 2014;8:329–333.
- Aldojain A, Jaradat J, Summersgill K, et al. Ectomesenchymal chondromyxoid tumor: a series of seven cases and review of the literature. *Head Neck Pathol.* 2015;9:315–322.
- Pires FR, Abrahao AC, Cabral MG, et al. Clinical, histological and immunohistochemical features of ectomesenchymal chondromyxoid tumor. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2009; 108:914–919.

- de Visscher JG, Kibbelaar RE, van der Waal I. Ectomesenchymal chondromyxoid tumor of the anterior tongue. Report of two cases. *Oral Oncol.* 2003;39:83–86.
- 21. Siegfried A, Romary C, Escudie F, et al. RREB1-MKL2 fusion in biphenotypic "oropharyngeal" sarcoma: new entity or part of the spectrum of biphenotypic sinonasal sarcomas? *Genes Chromosomes Cancer*. 2017;57:203–210.
- Nikitakis NG, Argyris P, Sklavounou A, et al. Oral myoepithelioma of soft tissue origin: report of a new case and literature review. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2010;110:e48–e51.
- Cen B, Selvaraj A, Prywes R. Myocardin/MKL family of SRF coactivators: key regulators of immediate early and muscle specific gene expression. *J Cell Biochem.* 2004;93:74–82.
- Selvaraj A, Prywes R. Megakaryoblastic leukemia-1/2, a transcriptional co-activator of serum response factor, is required for skeletal myogenic differentiation. J Biol Chem. 2003;278:41977–41987.
- Mokalled MH, Johnson A, Kim Y, et al. Myocardin-related transcription factors regulate the Cdk5/Pctairel kinase cascade to control neurite outgrowth, neuronal migration and brain development. *Development*. 2010;137:2365–2374.
- Huang D, Sumegi J, Dal Cin P, et al. C11orf95-MKL2 is the resulting fusion oncogene of t(11;16)(q13;p13) in chondroid lipoma. *Genes Chromosomes Cancer*. 2010;49:810–818.
- Flucke U, Tops BB, de Saint Aubain Somerhausen N, et al. Presence of C11orf95-MKL2 fusion is a consistent finding in chondroid lipomas: a study of eight cases. *Histopathology*. 2013;62:925–930.
- Garcia CB, Shaffer CM, Alfaro MP, et al. Reprogramming of mesenchymal stem cells by the synovial sarcoma-associated oncogene SYT-SSX2. Oncogene. 2012;31:2323–2334.